

Journal of the Algerian Chemical Society

Journal homepage: https://www.jacs-dz.org

ISSN 1111-4797



HPLC Chiral Analysis and Stereo-Bioactivity of Repaglinide Enantiomers

Djawhara Haddad, Nasser Belboukhari, Khaled Sekkoum, Meriem Bouanini, Marwa Ghouizi

Bioactive Molecules and Chiral Separation Laboratory, Faculty of exact sciences, Tahri Mohammed University, Istiklal street PO 417 Bechar, 08000, Algeria

Article History

Received:28/12/2024

Revised: 19/05/2025

Accepted: 26/05/2025

ABSTRACT

In this study a rapid chiral High performance Liquid Chromatography (HPLC) method has been developed and validated for the enantiomeric separation of Repaglinide, with the molecular docking study of Repaglinide against carbonic anhydrase II (CA2). The method was carried out on a chiralcel® OD-RH column using a mixture of n-hexane: 2-propanol (95:5, v/v), The flow rate was 1.0 ml/min and the eluent was monitored at 240nm. The software used in this molecular docking is PyRx-Virtual Screening Tools (for molecular docking process) and Discovery Studio (for pose visualization and data analysis). According to the study's findings, for HPLC two peaks with a 2.074- and 3,258-min retention time was achieved and the resolution, capacity and selectivity factors obtained were Rs = 0.773; k'1= 1.074, k'2 = 2.258 respectively and $\alpha = 2,1$. The result of molecular docking showed the binding energy of repaglinide to CA2 was found to be -6.8 kcal/mol with RMSD of 1.55 Å, indicating a higher binding affinity compared to native ligand (dansylamide). The method was found to be fast, simple, precise and suitable for analysis of Repaglinide in drug substance.

Keywords: *Repaglinide*, HPLC, Chiralcel[®]OD-RH, molecular docking, antidiabetic, chiral separation.

1. INTRODUCTION

Diabetes mellitus (DM) is one of the oldest diseases that has recently gained popularity and become a serious health concern for human [1,2]. It is a long-term metabolic condition defined by pancreatic β-cell dysfunction, decreased insulin production and resistance [3,4], with insulin being the major hormone directing intermediate metabolism [5]. Diabetes mostly affects the liver and kidney. Type 2 diabetes (non-insulin-dependent) can now be treated with orally administered hypoglycemic medications that help to lower blood sugar levels [1,2]. Carbonic anhydrase II (CAII) is a zinc metalloenzyme that is involved in numerous physiological processes in the human body, including pH regulation, electrolyte balance, ionic transport, carboxylation or decarboxylation reactions, biocalcification, and tumorigenesis. It catalyzes the reversible conversion of CO2 into bicarbonate, which is required for respiration, pH regulation, ion exchange, and bone resorption [6-9].

Repaglinide is an oral meglitinide-class antidiabetic medication used to treat type II diabetes mellitus. It reduces blood glucose levels by increasing insulin secretion from the pancreas by inhibiting the sulfonyl urea receptor on KATP channels in pancreatic β-cell membranes [10-14]. Repaglinide structure, binding profile, duration of action, and excretion method set it apart from other antidiabetic drugs because it is easily absorbed from the gastrointestinal tract after oral administration. [15,16]. Chemically, Repaglinide is a novel carbomoxylmethyl benzoic acid derivative, also referred to as (S)-2-ethoxy 4-[[3-methyl-1-[2-(1-piperidinyl) phenyl]butyl]amino]-2-oxoethyl][17-19], as illustrated in Figure 1. Repaglinide has the molecular formula C₂₇H₃₆N₂O₄ and is soluble in methanol and methylene chloride but practically insoluble in water [20].

Over several decades, researchers have grown increasingly interested in and focused on the separation of chiral enantiomers of medications [21,22]. In the field of medicine, pharmacology, and chemical research, determining enantio-separation, absolute configuration, and chiral recognition mechanism is crucial for the production of safe and efficient medicines during the drug development process. Chiral medications enantiomers may differ in their biological activity as well as in their potential therapeutic effects [23-25]. Several methods have been reported in the literature for determination of Repaglinide in pharmaceutical formulations and in human serum and other biological fluids, these methods include HPLC [26-29], HPLC/ tandem mass spectrometry [11,30,31], visible spectrophotometric [32-34], HPTLC [35-36] and electrochemical methods [16,37]. HPLC is the most commonly used method for chiral analysis because it offers greater

resolution, sensitivity, broad applicability, and compatibility with a wide range of detection techniques. It is crucial to determine the validity, pharmacokinetics, and pharmacodynamics of chiral medications at various stages of drug development [23,24].

The literature provides an HPLC method for the enantio-separation of *Repaglinide* using a Chiralpack®AD-H (amylose-based stationary phase) column [26], Chiralpack®IA (immobilized amylose based stationary phase) [27]and C18 column [10-20, 28-35].

Molecular docking is a widely used tool in drug discovery and development to investigate protein-ligand interaction configurations [38-40]. This method entails docking a molecule with a certain macromolecule and determining the binding free energy between the ligand and receptor. Molecular docking can save time, effort, and environmentally harmful solvents or chemicals [6,41]. The current study addresses with a fast, effortless, precise, robust enantioselective, and isocratic chiral LC technique for the enantio-separation of *Repaglinide* employing a Cellulose tris (3, 5 Dimethylphenyl carbamate) based chiral stationary phase (chiralcel®OD-RH) (Fig.2) and Molecular docking simulations were done using PyRx and Discovery Studio software to assess the binding affinity and mechanism of interaction between *Repaglinide* and the CA2 enzyme (Fig.3). The availability of highly sensitive and selective HPLC technology will be very useful in the determination of *Repaglinide* in pharmaceutical dosage forms.

Fig.1. Chemical structure of Repaglinide Fig.2. Chemical structure of chiralcel[®]OD-RH column

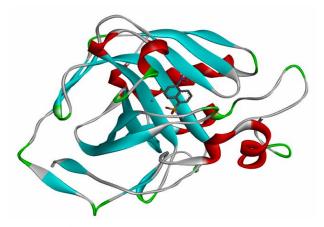


Fig.3. 3D structure of CAII

2. RESULTS AND DISCUSSION

2.1. Chiral Separation Study:

The mechanism of separation in direct chiral separation methods is the interaction of chiral stationary phase (CSP) with enantiomers that are analytes to form short-lived, transient diastereomeric complexes [26] [42,43]. Chiral separation of *Repaglinide* can be achieved using chiral stationary phases (CSPs) such as chiralcel®OD-RH column which is Cellulose tris (3, 5-dimethylphenylcarbamate) coated on 5µm silica-gel. The Cellulose-based stationary phase in Chiralcel®OD-RH column has higher selectivity being suitable for the enantioselective separation and accurate quantification of (R)-*Repaglinide*. The results of the chiral liquid chromatographic method are presented in table 1.

Table 1: Result of chiral analysis of *Repaglinide* by HPLC on chiralcel[®]OD-RH column

| Csp | Peak | Rt (Rt) | K' | α | Rs | % |
|-----------------|------|---------|-------|-----|-------|----|
| Chiralcel®OD-RH | 1 | 2.074 | 1.074 | | | 13 |
| | 2 | 3,258 | 2.258 | 2.1 | 0.773 | 87 |

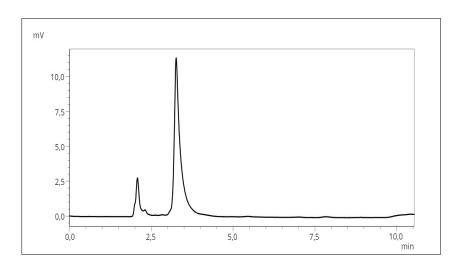


Fig.4. A Typical HPLC chromatogram of Repaglinide on chiralcel®OD-RH column. Mobile phase consisted of n-hexane: 2-propanol (95:5v/v), flow rate 1.0 mL min, UV-240 nm; column ambient temperature

A representative chromatogram of the enantiomeric resolution of *Repaglinide* is shown in figure 4. In chiralcel®OD-RH, the retention times of Repaglinide were found at 2.074 and 3.258 min. A good resolution (Rs = 0.773) between two enantiomers was obtained, with adequate system suitability parameters (enantioselectivity α= 2.1, Capacity Factor K₁=1.074 and K₂=2.258). In C18 column, the retention time of *Repaglinide* was found at 1.95 min.Chiral separation of *Repaglinide* using Chiralcel®OD-RH column is an efficient way to isolate the two enantiomers of drug. Following the results obtained, chiral separation of *Repaglinide* using a Chiralcel®OD-RH column was an effective method to isolate the two optical isomers of the drug.

1. Molecular docking study

Molecular docking study was performed to investigate the binding affinity of *Repaglinide* and dansalymide to CII enzyme. The obtained RMSD results were good with a value of less than 2 Å, indicating accurate prediction according to the literature [44–46]. Table 2 summarizes the binding energy of the study (kcal/mol), RMSD values (Å) and type of interaction with 1OKL.

Table 2. The results of molecular docking

| Compounds | Binding Energy | RMSD | Interactions residues and type interaction |
|---------------|-----------------|-------|--|
| Native ligand | | 1.95Å | hydrophobic pi alkyl interactions (VAL121, PHE |
| | -5.5 Kcal /mol | | 130, LEU197) |
| | | | Pi sigma (LEU197) |
| | | | Pi sulfur (THR198) |
| | | | Hydrogen bond (LEU197, THR198) |
| S-repaglinide | | 1.55Å | pi-pi T-shaped (TRP244, PHE230) |
| | | | pi alkyl (PRO13, LYS9) |
| | -6.8 Kcal /mol | | attractive charge (ASP242) |
| | -0.6 Kcai/IIIoi | | unfavorable negative-negative (GLU238) |
| | | | Hydrogen bonds (THR7, LEU239) |
| | | | Pi sigma (PHE230) |

Table 2 indicates that *Repaglinide* has a low affinity binding value of -6.8 kcal / mol compared to dansalymid -5.5 kcal / mol has a binding value, with respective RMSD values of 1.55 and 1.95 Å.

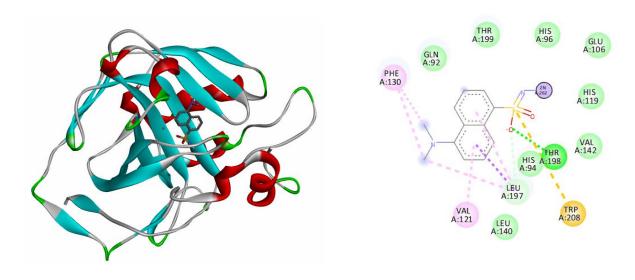


Fig.5. 3D and 2D docked pose depiction of dansylamide

The native ligand (dansylamide) formed hydrophobic pi alkyl interactions with VAL121, PHE130 and LEU197. Pi sigma interaction was constructed from the benzene ring to LEU197 amino acid, while pi sulphur interaction binding was achieved with THR198. The LEU197 and THR198 residues formed H bonds towards oxygen atom in the dansylamide molecule (Figure 5).

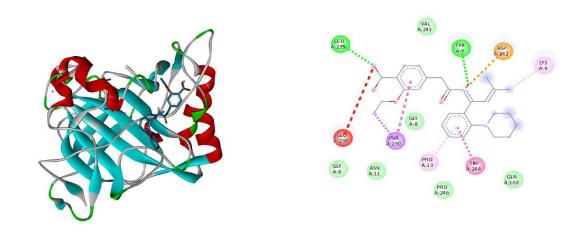


Fig.6. 3D and 2D docked pose depiction of Repaglinide

When the interaction of *Repaglinide* with the residues in the enzyme's active site was examined, it was observed that it made two different hydrogen bond interactions, two different pi alkyl interactions, pi sigma interaction, two different pi-pi T-shaped interactions, attractive charge and unfavorable negative-negative interaction. Two different pi-pi T-shaped interaction were constructed from the benzene ring to TRP244 and PHE230. Of the hydrophobic interactions formed, pi alkyl interactions were achieved with PRO13 and LYS9 residues, while attractive charge interaction was achieved with ASP242 and unfavorable negative-negative interaction formed with GLU238 amino acid. Accordingly, all H-bonds formed are from the S-*Repaglinide* molecule via amino acids THR7 and LEU239 and Pi sigma interaction formed with PHE230 (Fig.6).

3. EXPERIMENTAL

3.1.Apparatus:

The HPLC system used was a SHIMADZU Scientific Instruments' system LC-20A (Shimadzu, Kyoto, Japan) with an injector of 10 µl Rheodyne 1907 sample loop, a pump LC-20A, a vacuum degasser DGU-20A5, a UV detector Shimadzu SPD-20A (Kyoto, Japan), Shim-pack GIS C18

column, and Chiralcel®OD-RH coated on a 5μm silica-gel column. The LC Lab solution software (Shimadzu, Tokyo, Japan) was used to obtain, store, and evaluate chromatographic data. The chromatographic separation was performed on two mobile phases: methanol for the Shim-pack GIS C18 column and n-hexane:2-propanol (95:05, v/v) for the Chiralcel®OD-RH. The mobile phases were pumped from the solvent reservoir to the column at a flow rate of 1 ml/min. The column temperature was maintained at ambient temperature, and the eluent was monitored at a wavelength of 240 nm. The injection volume was 10 μL.

3.2.Chemicals:

Novo Norm[®] tablets: Each tablet contained 2 mg *Repaglinide*. Chromasolv[®]methanol, LiChrosolv[®]n-hexane and Chromasolv[®] 2-propanol for HPLC were purchased from Sigma Aldrich. The stock solution of *Repaglinide* (2 mg) was prepared by dissolving appropriate amount of substance in methanol.

3.3. Molecular Docking Methodology

Ligands preparation: Three-dimensional structure of S repaglinide and native ligand (dansylamide), downloaded from the site of databank zinc.docking.org in the form of mol2 code ZINC22056467 and mol2 code ZINC56543 respectively. The software utilized is PyRx-AutoDock Vina and Discovery Studio.

Protein preparation: The material comprises a three-dimensional structure of Carbonic anhydrase II (CAII) with the code of 1okl, which is downloaded from the website of protein data bank https://www.rcsb.org/structure/1OKL. The CAIIprotein is separated by water and ligand molecules with Discovery Studio version 4.5. Then, it is stored in a pdb format. Next, with Autodock Vina 4.2, the hydrogen atom of the protein is displayed and stored in a pdbqt format.

Molecular docking: After preparing the ligands and Protein, PyRx was implied with the Autodock Vina option using the new scoring function. It analyzes the docking orientations and interactions between the ligands and CAII. Determination of Grid box of all ligands is at the receptor site (CAII) with the centre X: -1.8654, Y: -2.6221, Z: 15.5964 and the large dimension with the angstrom X: 54.305, Y: 55.3329, Z: 66.9195 for molecular docking. After importing the docked data into the Discovery Studio visualizer, 2D and 3D formats of the significant interactions between the ligands and the receptor-binding sites were obtained.

4. CONCLUSION

The results of HPLC method showed that it is a flexible, simple and fast method for the chiral separation and detection of enantiomers in a pharmaceutical study. Chiralcel®OD-RH was found to be selective for the enantiomers of the drug. The mobile phase is simple to prepare and economical. The developed method is more rapid and enantioselective than reported methods. Hence, this can be conveniently adopted for routine analysis of *Repaglinide* in pure form and its dosage forms. The results of molecular docking using PyRx shows that *Repaglinide* has a more stable interaction with carbonic anhydrase II compared to *Dansylamide* based on its lower binding energies. This enables the production of highly pure and effective drugs, minimizing side effects and ensuring consistency in dosing.

5. REFERENCES

- [1] Penta,J.; Gorre,T.; Reddy, Y. N.pharmacokinetic and pharmacodynamic interaction study of curcumin with repaglinide in normal and diabetic rats.*J. Global Trends Pharm Sci.* 2017, 8, 3, 4130 4137.
- [2] Yao,J.; Shi,Y. Q.;Zhuo, Z. R. Li; Jin,S. H.Development of a RP-HPLC method for screening potentially counterfeit anti-diabetic drugs. *J. Chromatogr. B. Analyt. Technol. Biomed.Life. Sci.* 2007,853,1, 54–259.
- [3] Dhole,S. M.; Amnerkar, N. D.; Khedekar,P. B.comparison of uv spectrophotometry and high-performance liquid chromatography methods for the determination of repaglinide in tablets. *Pharm Methods*.2012, 3, 2, 68–72.
- [4] Uppu,P.; Kandukoori,N.; N. Yellu. effect of allicin on pharmacokinetics and pharmacodynamics of repaglinide in normal and streptozocin induced diabetic rats. *Int J Biol Pharm Allied Sci*.2021,10, 8,2571-2577.
- [5] Sonia, K.; Nappinnai, M.; Manikandan, K. stability indicating rp-hplc method for the estimation of metformin hydrochloride and repaglinide as api and estimation in tablet dosage form. Int. J. Pharma. Quality Assurance .2016, 7,3, 46-50.
- [6] Aditama, R.; Mujahidin, D.; Syah, Y. M.; Hertadi, R. docking and molecular dynamics simulation of carbonic anhydrase ii inhibitors from phenolic and flavonoid group. *Procedia Chem.* 2015,16,357–364.
- [7] Sağlık, B. N. *et al.* synthesis, molecular docking analysis and carbonic anhydrase i-ii inhibitory evaluation of new sulfonamide derivatives. *Bioorg Chem.* 2019, 91,2,103153.

- [8] Sharker, M. R.; Sukhan, Z. P.; Sumi, K. R.; Choi, S. K. K. S.; Kho, K. H. molecular characterization of carbonic anhydrase ii (ca ii) and its potential involvement in regulating shell formation in the pacific abalone, haliotis discus hannai. *Front Mol Biosci.* 2021, 7, 8,669235.
- [9] Jakubowski, M.; Szahidewicz, E. K; Doroszko, A. the human carbonic anhydrase ii in platelets: an underestimated field of its activity. *BioMed Research International*. 2018, 28, 4548353.
- [10] El-Zaher, A. A; Elkady, E. F.; Elwy, H. H.; Saleh, M.A. validated liquid chromatographic method for simultaneous determination of metformin, pioglitazone, sitagliptin, repaglinide, glibenclamide and gliclazide application for counterfeit drug analysis. *J Anal Bioanal Tech*, 2015, S13.
- [11] Zhang,J.; Gao,F.; Guan,X.; Sun,Y.; Gu,J.; Paul Fawcett,J. determination of repaglinide in human plasma by high-performance liquid chromatography—tandem mass spectrometry. *Acta Pharm Sin B*.2011, 1, 1, 40–45.
- [12] Patel, D. R.; Patel, L. J.; Patel, M. M. development and validation of stability indicating method for the determination of repaglinide in pharmaceutical dosage form using high performance liquid chromatography. *Int. J. ChemTech Res.* 2011, 3, 2, 539-546.
- [13] Tatiparthi,R.; Bannoth,C. K.method development and validation of metformin and repaglinide in rabbit plasma by rp-hplc. *FABAD J. Pharm. Sci.* 2010, 35, 69-75.
- [14] D. G. Han *et al.*, "Pharmacokinetic evaluation of metabolic drug interactions between repaglinide and celecoxib by a bioanalytical HPLC method for their simultaneous determination with fluorescence detection," *Pharmaceutics*, 2019, 11, 8, 382.
- [15] Prashant, P.; Nandasana, P.V. Development and Validation of RP-HPLC Method for the Estimation of Repaglinide in Bulk Drug and Pharmaceutical Formulation. *Int. J. Drug. Dev.Res* .2012, 4,3,247-252.
- [16] Abdel Nabi, M. E.; Genidy, G.M.; ATTIA, K. electrochemical determination of the antidiabetic drug repaglinide. *Pharmaceutical Society of Japan*. 2008, 128, 1,171-7.
- [17] Ruzilawati, A. B.; Wahab, M. S. A.; Imran, A.; Ismail, Z.; Gan, S. H. Method development and validation of repaglinide in human plasma by HPLC and its application in pharmacokinetic studies," *J Pharm Biomed Anal.* 2007, 43, 5,1831–1835.
- [18] Prameela, A. R; Sekaran, B. C.; Teja, S. P. "determination of repaglinide in pharmaceutical formulations by rp-hplc method. Journal of Applied Sciences Research. 2009, 5,10,1500-1504.

- [19] Navamanisubramanian, R.; Panchagiri, S.; Nerella, R.; Duraipandian, C.; Seetharaman, S.stability indicating rp-hplc method for estimation of repaglinide in rabbit plasma. *International Journal of Applied Pharmaceutics*. 2019,11,3,206–210.
- [20] Ramakrishna,B.;Mondal,S. a review of analytical methods for determination of type-ii antidiabetic drugs in pharmaceuticals and biological matrices. *Asian. J. Pharm. Clin. Res.* 2021, 14, 1, 69-76.
- [21] IbrahimA. E.; *et al.* recent advances in chiral selectors immobilization and chiral mobile phase additives in liquid chromatographic enantio-separations: a review. *Journal of Chromatography A*, 2023,1706,464214.
- [22] Rahou, I.; Sekkoum, K.; Belboukhari, N.; Cheriti, A.; Aboul-Enein, H. Y. liquid chromatographic separation of novel 4-amino-flavanes series diastereomers on a polysaccharide-type chiral stationary phase. *J Chromatogr Sci*.2016, 54, 10, 1787–1793.
- [23] Imran, A. *et al.* determination of enantio-separation, absolute configuration and chiral recognition mechanism of ofloxacin and flumequine by hplc and modeling studies," *Journal of Chemical Technology and Biotechnology*. 2021, 96,10, 2901–2908.
- [24] Al-Sulaimi,S.; Kushwah,R.;Abdullah,M.A.; El Jery,A.;Aldrdery,M.; Ashraf,G. A. emerging developments in separation techniques and analysis of chiral pharmaceuticals. *Molecules* .2023, 28, 17, 6175.
- [25] Bounoua, N.; Sekkoum, K.; Belboukhari, N.; Cheriti, A.; Aboul-Enein, H. Y. achiral and chiral separation and analysis of antifungal drugs by hplc and ce: a comparative study: mini review. *Journal of Liquid Chromatography and Related Technologies*. 2016, 39, 11, 513–519.
- [26] Patil,K.; Rane,V.;Yeole,R.; Shinde,D.a validated chiral lc method for the enantiomeric separation of repaglinide on immobilized amylose based stationary phase. *J. Braz. Chem. Soc.* 2012, 23, 6, 1048-1053.
- [27] Rane, V. P.; Shinde, D. B. a validated chiral lc method for the enantiomeric separation of repaglinide on amylose based stationary phase," *Chromatographia*. 2007, 66, 7–8,583–587.
- [28] Chandra, M.S.; Sharma, S. stability indicating rp-hplc method for determination and validation of repaglinide in pharmaceutical dosage form. *Int.J. Chem. Tech. Res.* 2011, 3, 1, 210-216.
- [29] Raja, A.; Santhoshi, R.; David, B.; Roa, K.N.V.; Selva, K.rp-hplc method development and validation for simultaneous estimation of metformin and repaglinide in bulk and tablet dosage form. *Asian journal of research in pharmaceutical science and biotechnology*. 2015, 3,2,33-40.

- [30] Emmie, Ho. N. M., *et al.* Detection of anti-diabetics in equine plasma and urine by liquid chromatography-tandem mass spectrometry. *Journal of Chromatography B.* 2004, 811,1,65–73.
- [31] Fayyad,M. K.;Ghanem,E. H. liquid chromatography tandem mass spectrometry method for determination of anti-diabetic drug repaglinide in human plasma. *Am J Analyt Chem*.2014, 05,04,281–290.
- [32] Pritosh, P.; Panda, O, P.; Chandan, M.; Podila, N.new rp-hplc method for the estimation of repaglinide in bulk and in pharmaceutical dosage forms. *Int J Pharma Sci.* 2013, 3, 2,189-193.
- [33] Aslan,S. S.; Yılmaz,B. derivative spectrophotometric and isocratic high performance liquid chromatographic methods for simultaneous determination of repaglinide and metformin hydrochloride in pharmaceutical preparations. *Am J Analyt Chem*, 2017, 8, 9,541–552.
- [34] Dash,A. K. method development, validation and stability study of repaglinide in bulk and pharmaceutical dosage form by uv spectrophotometric method. *International Journal of Biological and Pharmaceutical Research*. 2011, 2, 1,7-10.
- [35] Abdelhamid, N. S.; Elsaady, M. T.; Ali, N. W.; Abuelazem, W. G. simultaneous determination of repaglinide, metformin hydrochloride and melamine by new hplc and hptlc chromatographic methods. *Analytical Chemistry Letters*, 2019, 9,3,418–429.
- [36] JiladiaM. A.; Pandya,S. S. estimation of repaglinide in bulk and tablet dosage forms by hptlc method. *International Journal of Pharmacy and Pharmaceutical Sciences*. 2009, 1, 11.
- [37] Pawar, S. K.; Jaldappagari, S. interaction of repaglinide with bovine serum albumin: spectroscopic and molecular docking approaches. *J Pharm Anal.* 2019, 9,4,274–283.
- [38] Mimouni, F. Z.; Belboukhari, N.; Sekkoum, K.; Aboul-Enein, H. Y.novel gatifloxacin3-carboxamide derivatives as anti-tumor agents: synthesis, enantioseparation, and molecular docking. *Curr Anal Chem.* 2022, 18, 10, 1108–1116.
- [39]Oulad Ali,H.; Belboukhari,N.; Sekkoum, K.;Belboukhari, M.;Seddiki,L.S. Computational molecular docking analysis of linalool enantiomers interaction with mitogen-activated protein kinase 1 (mapk1): insights into potential binding mechanisms and affinity. *Chirality*. 2025, 37, 3.
- [40] Tajiani,F.; Ahmadi,S.; Lotfi,S.; Kumar, P.; Almasirad,A. in-silico activity prediction and docking studies of some flavonol derivatives as anti-prostate cancer agents based on monte carlo optimization. *BMC Chem*.2023, 17, 1.

- [41] Hamache, T.; Belboukhari, N.; Sekkoum, K.; Zaid, M. E. A. evaluating the therapeutic potential of 4-hydroxyflavanes diastereomers derivatives against (MetAP2) for anti-cancer therapy: a molecular docking study. *Chemical Product and Process Modeling*. 2025, 2024-0098.
- [42] Rahou, I.; Belboukhari, N.; Sekkoum, K.; Cheriti, A.; Aboul-EneinH. Y. chiral separation of 4-iminoflavan derivatives on several polysaccharide-based chiral stationary phases by hplc," *Chromatographia*. 2014, 77, 17–18, 1195–1201.
- [43] Bouanini, M.; Belboukhari, N.; Sekkoum, K.; Haddad, D. Synthesis, Enantio-Separation and Molecular Docking Study of Hesperidin-Oxime. *Egyptian Journal of Chemistry*. 2025, 68, 5, 401 408.
- [44] Widiyarti, G.; Firdayani, M.; Hanafi, M.; Kosela, S.; Budianto, E. molecular docking of citronellol, geraniol and ester derivatives as pim 1 kinase inhibitor of leukemia cancer. *Jurnal Kimia Valensi*. 2019, 5, 2,133–142.
- [45] Belal,A. *et al.*design of new captopril mimics as promising ace inhibitors: adme, pharmacophore, molecular docking and dynamics simulation with mm-pbsa and pca calculations. *Journal of Taibah*.2023,17, 1.
- [46] Rahmadania, N. H.;R. Trijuliamos, M. molecular docking of turmeric active compounds (curcuma longa 1.) against main protease in covid-19 disease. *East Asian Journal of Multidisciplinary Research (EAJMR)*, 1,3,353–364.